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Chapter 1 : A Photographic Atlas for the Microbiology Laboratory - just-pdf

The third edition of the Photographic Atlas for the Microbiology Laboratory is one of the best selling microbiology books in the higher education market. The authors have built on the success of this book by making significant improvements for the new edition.

Unknown Report includes hypotheses and references 25 Notebook: Assignments more than one week late will not be graded. Required Lab Supplies Lab coat There is a limited supply of lab coats in the lab. Lab coats can also be purchased at the bookstore. Safety glasses supplied in lab Optional Lab Supplies 3 x 5 cards a file box is handy to keep your cards in Absentee Policy Attendance in lab is mandatory. Only University excused absences will be allowed. You must contact your TA or Professor prior to missing the lab and show appropriate paperwork upon your return. Unexcused absences will not be accepted. One unexcused absence will result in the loss of 5 points and two unexcused absences will result in an automatic F in the course. Missed labs must be made up during open lab hours. Not only does this allow you to be efficient and productive in the time allowed for the lab, it facilitates a better understanding of the material. Many of the exercises performed in this lab require several days to complete. Generally, each lab period will involve finishing the previous exercise and then starting another. At times this can be a little chaotic. Therefore, it is imperative that you concentrate on completing each lab by recording all of the results especially for large group projects , and answering the questions for each lab before starting with the next lab exercise. This course covers a good number of specific microorganisms, microbiological tests, and procedures; the index cards may be an invaluable study aid. Laboratory Safety Lab coats and safety glasses must be worn at all times while in the laboratory. Regular prescription glasses are generally not made with safety glass and thus safety glasses must be worn over the top of these glasses at all times. Safety glass checks will be made throughout the semester and students without their glasses will loose 2 points. All other items such as coats, books, and bags should be stored on the shelves provided for this purpose. No eating, drinking or smoking in lab. Know lab safety procedures and the location of the first aid kit, eyewash, and fire extinguisher. All culture material should be handled as if it were potentially harmful. Be very careful with Bunsen burners. Burners should be turned off when not in use. Long hair must be tied back at all times while in the laboratory. Dispose of materials as instructed. Do not carelessly throw materials in wastebaskets or sinks; biohazard waste containers are available. Report any accident or injury immediately to the laboratory instructor so that prompt action can be taken. After each lab, WASH your hands before leaving the laboratory.

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Chapter 2 : [PDF] Download A Photographic Atlas For The Microbiology Laboratory Free | Unquote Books

Photographic Atlas for the Microbiology Laboratory / Edition 4 This full-color atlas is intended to act as a supplement to introductory microbiology laboratory manuals. It is not designed to replace them, nor is it intended to replace actual performance of the techniques.

Today, more sophisticated compound light microscopes are routinely used in microbiology laboratories. The various types of light microscopy include bright-field, dark-field, fluorescence, and phase contrast microscopy. Although each method has specific applications and advantages, bright-field microscopy is most commonly used in introductory classes and clinical laboratories. Many research applications use electron microscopy because of its ability to produce higher quality images of greater magnification. Light Microscopes

Bright-field microscopy produces an image made from light that is transmitted through a specimen (Figure A). Because most biological specimens are transparent, contrast between the specimen and the background can be improved with the application of stains to the specimen (see Sections 5 and 6). The price of improved contrast is that the staining process usually kills cells. This is especially true of bacterial staining protocols. Image formation begins with light coming from an internal or an external light source (Figure). It passes through the condenser lens, which concentrates the light and makes illumination of the specimen more uniform. Because of its thickness, the entire organism will not be in focus at once. Continually adjusting the fine focus to clearly observe different levels of the organism will give a sense of its three-dimensional structure. The bright rods are bacteria. B The same diatom viewed with dark-field microscopy. Notice also that the bacteria are not visible, though this would not always be the case. C This phase contrast image of the same diatom shows different details of the interior than what is seen in the other two micrographs. Also, notice the bacteria are dark. D This is a fluorescence micrograph of *Mycobacterium kansasii*. The apple green is one of the characteristic colors of fluorescence microscopy. It then enters the objective lens, where it is magnified to produce a real image. The real image is magnified again by the ocular lens to produce a virtual image that is seen by the eye through the objective lens from the specimen produces a magnified real image. This image is magnified again as it passes through the ocular lens to produce a virtual image that appears below or within the microscope. The amount of magnification produced by each lens is marked on the lens (Figure A and B). Total magnification of the specimen can be calculated by using the following formula: Plan means the lens produces a flat field of view. Apochromatic lenses are made in such a way that chromatic aberration is reduced to a minimum. From left to right, the lenses magnify 10X, 20X, and 40X, and have numerical apertures of 0. The 20X lens has other markings on it. The mechanical tube length is the distance from the nosepiece to the ocular and is usually between 160 to 175 mm. However, this 20X lens has been corrected so the light rays are made parallel, effectively creating an infinitely long mechanical tube length. This allows insertion of accessories into the light path without decreasing image quality. The thickness of cover glass to be used is also given (0.18 mm). B A 10X ocular lens. C A condenser removed from the microscope with numerical aperture of 1. The lever at the right is used to open and close the iris diaphragm and adjust the amount of light entering the specimen. The practical limit to magnification with a light microscope is around 1000X. Although higher magnifications are possible, image clarity is more difficult to maintain as the magnification increases. Clarity of an image is called resolution (Figure). The limit of resolution or resolving power is an actual measurement of how far apart two points must be in order for the microscope to view them as being separate. Notice that resolution improves as resolving power is made smaller. The best limit of resolution achieved by a light microscope is about 0.2 μm . That is, at its absolute best, a light microscope cannot distinguish between two points closer together than 0.2 μm . For a specific microscope, the actual limit of resolution can be calculated with the following formula: Because numerical aperture has no units, the units for D are the same as the units for wavelength, which typically are in nanometers (nm). As you look at the cars in the foreground of the photo, it is easy to see both headlights as

separate objects. When the apparent distance between automobile headlights reaches about 0. Using immersion oil between the specimen and the objective lens increases its numerical aperture and in turn, makes its limit of resolution smaller. If necessary, oil may also be placed between the condenser lens and the slide. The result is better resolution. The light microscope may be modified to improve its ability to produce images with contrast without staining, which often distorts or kills the specimen. In dark-field microscopy Figure B , a special condenser is used so only the light reflected off of the specimen enters the objective. The appearance is of a brightly lit specimen against a dark background, and often with better resolution than that of the bright field microscope. Phase contrast microscopy Figure C uses special optical components to exploit subtle differences in the refractive indices of water and cytoplasmic components to produce contrast. Light waves that are in phase that is, their peaks and valleys exactly coincide reinforce one another and their total intensity because of the summed amplitudes increases. Light waves that are out of phase by exactly one-half wavelength cancel each other and result in no intensity—that is, darkness. Wavelengths that are out of phase by any amount will produce some degree of cancellation and result in brightness that is less than maximum but more than darkness. Thus, contrast is provided by differences in light intensity that result from differences in refractive indices in parts of the specimen that put light waves more or less out of phase. As a result, the specimen and its parts appear as various levels of darks and lights. Fluorescence microscopy Figure D uses a fluorescent dye on the specimen that emits fluorescence when illuminated with ultraviolet radiation. In some cases, specimens possess naturally fluorescing chemicals and no dye is needed. The Electron Microscope The electron microscope uses an electron beam to create an image, with electromagnets acting as lenses. The limit of resolution is improved by a factor of theoretically down to 0. The transmission electron microscope TEM Figure produces a two-dimensional image of an ultrathin section by capturing electrons that have passed through the specimen. The degree of interaction between the electrons and the heavy metal stain affects the kinetic energy of the electrons, which are collected by a fluorescent plate. A sample transmission electron micrograph is shown in Figure The previous paragraph gave a brief overview of how the TEM works. However, a key to successful transmission electron microscopy is excellent sample preparation. Following is an overview of sample preparation. The specimen is fixed by one of various methods treatment with formaldehyde, glutaraldehyde, or osmium tetroxide to prevent cell decomposition, stained with an electron dense material lead, uranium, or osmium compounds , dehydrated, and embedded in a plastic block Figure It is then cut into thin slices using an ultramicrotome Figure armed with a glass or diamond blade. The image is then viewed on the monitor. Since light is not used, the image is not in color. These cells were magnified 12,X Figure , which is inserted into the TEM so it rests in the electron beam path. Figure shows what the microscopist sees when working. On the right is a trimmed block that has had excess resin cut away to produce a minute piece of specimen that extends from the block. This is the portion of specimen to be sectioned Figure The arm holding the specimen traces an elliptical path as it approaches and is withdrawn from the sample. In each cycle, it is advanced the distance equal to the desired section thickness, often nm. B The specimen block S , with the tiny, trimmed down specimen arrow facing the blade B1 , is held in the ultramicrotome chuck. As the specimen moves forward and passes by the glass or diamond blade a thin slice is made, which is caught and floated on water in the boat Bt behind the blade. The specimen holder is withdrawn, returned to the starting position, and advanced by the desired thickness and another cut is made. The process is repeated to produce multiple sections of the same thickness. The shiny material between grid bars is a plastic film that fills in the openings and keeps specimens from dropping through. The grid is placed in a grid holder that is inserted into the TEM. In order for the image to be seen, the microscopist views the specimen on this screen coated with a phosphorescent material. The kinetic energy of the electrons hitting the screen is converted to light, which makes the specimen visible. The thick, dark lines are the grid bars at very low magnification. The image is also captured by a digital camera and viewed on a computer monitor. Some electrons are reflected backscatter electrons , whereas other electrons secondary electrons are emitted from the metallic stain. These electrons are captured and used to produce the three-dimensional image. A sample

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scanning electron micrograph is shown in Figure . As with the TEM, sample preparation involves fixation, dehydration, and staining but not sectioning. Argon gas is ionized in an electric field within an evacuated chamber. The positively charged argon ions bombard a gold foil, which releases gold atoms that are free to coat the sample. Figure shows a sputter coater. This micrograph is of E. Sputtering with gold occurs when ionized argon gas bombards a gold foil to release gold atoms. Two specimens are visible within; the purple is the argon gas. Suggested Reading - Michael J.

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Chapter 3 : Download [PDF] Microbiology Laboratory Theory And Application Free Online | New Books in F

Book Preface. As my fingers hit the keys of my laptop, I realize that I am, after a long, seemingly endless process, within days of completing the fourth edition of A Photographic Atlas for the Microbiology Laboratory.

Growth Patterns on Agar, Bacterial Growth, Atlas of Growth Patterns on Agar, A Photographic Atlas for the Microbiology Laboratory Purpose Recognizing different bacterial growth morphologies on agar plates is a useful and often crucial step in the identification process. Agar slants are typically used for cultivation of pure cultures. Bacteria also frequently display distinct morphological color and texture on agar slants. Principle When a single bacterial cell is deposited on a solid nutrient medium, it begins to divide. One cell makes two, two make four, four make eight. Eventually a colony appears where the original cell was deposited. Once the purity of a colony has been confirmed by an appropriate staining procedure Sections 5 and 6 , cells can then be transferred to a sterile medium to begin a pure culture. Color, size, shape, and texture of microbial growth are determined by the genetic makeup of the organism. However, organismal genetic expression is also greatly influenced by environmental factors including nutrient availability, temperature, and incubation time. Colony characteristics may be viewed with the naked eye or with the assistance of a colony counter Figure The basic categories of growth include colony shape, margin edge , elevation, color, and texture Figure Colony shape may be described as circular, irregular, or punctiform tiny. The margin may be entire smooth, with no irregularities , undulate wavy , lobate lobed , filamentous, or rhizoid branched like roots. Colony elevations include flat, raised, convex, pulvinate very convex , and umbonate raised in the center. Colony texture may be moist, mucoid, or dry. Pigment production is another useful characteristic and may be combined with optical properties such as opaque, translucent, shiny, or dull. The transmitted light and magnifying glass allow observation of greater detail, however, colony color is best determined with reflected light. The grid in the background is used as a counting aid. Each big square is a square centimeter. Descriptions also should include color, surface characteristics dull or shiny , consistency dry, butyrous-buttery, or moist and optical properties opaque or translucent. Figures through show a variety of bacterial colony forms and characteristics. Where applicable, contrasting environmental factors are indicated. Figures and show growth characteristics on agar slants. They range in size from 1 to 3 mm. They are about 1 mm in diameter. Rhodococcus species are soil organisms. Also note the extensions of growth along the streak line. They vary in size from 2 to 6 mm. While it is a normal inhabitant of the human intestinal tract, it is associated with community-acquired pneumonia and nososomal urinary tract infections. Note the raised center. B Close-up of the A. This species is also able to partially hemolyze red blood cells -hemolytic , as evidenced by the greening around each colony. They are translucent at the edges and about 5 mm in diameter. This species is highly resistant to ionizing radiation. B Close-up of circular C. The one on the right is getting there. Their diameters are about 3 mm. At a later stage of development, they produce yellow reproductive spores. This one plate fragranced the entire incubator! This is a photograph of P. It was almost liquid in composition, something that is indicated by its contact with the yellow colony to its right. It is found in soil and water, and rarely produces infections in humans. Chromobacterium violaceum produces a much more intense purple pigment when grown on Trypticase Soy Agar left than when grown on Nutrient Agar, a less nutritious medium right. B The same plate of S. B Close-up of B. Note the wormlike appearance. B Note the-hemolysis darkening of the agar; see page 61 for more information shown by much of the growth. C This is a close-up of the same plate as in B. Note the weak Beta-hemolysis of the white colony in the upper right arrow. White growth with Beta-hemolysis is characteristic of Staphylococcus aureus. From left to right: Suggested Reading - Michael J.

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Chapter 4 : {PDF} A Photographic Atlas For The Microbiology Laboratory || Free Download and Read eBook

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Ads Book Preface As my fingers hit the keys of my laptop, I realize that I am, after a long, seemingly endless process, within days of completing the fourth edition of A Photographic Atlas for the Microbiology Laboratory. Or so I thought. But, PAML 4e has presented its share of challenges. First, and foremost, is the fact that this is the first project out of three previous Atlas editions, three editions of Microbiology Laboratory Theory and Application, one edition of Microbiology Laboratory Theory and Applicationâ€”Brief Edition, and three editions of Exercises for the Laboratory Manual I have worked on without my longtime friend and colleague, Burt Pierce. Burt made the courageous and healthy decision to retire and move to Portland, OR, to enjoy life with his wife, three dogs, and two cats notice the absence of any microbes in the family. Yet, while he was not an active participant, his influence remains in this edition. A majority of his written and photographic contributions are still here, and I have tried to live up to his eye for detail, his demand for excellence, and his dedication to knowing our readership. Our skills complemented one another and the books were clearly better because of it. When we did the first edition, the Atlas broke the mold at Morton Publishing by including much more explanatory text for the photos beyond captions. In fact, the captions were criticized for not being very informative! While we have continued to add photos and expand coverage in each edition, the increase in text has considerably outpaced the photos. For those of you who have been with us through all four editions, compare photograph sizes in the first edition to this one! Writing is not easy for me. Chalk up another challenge being met. In many ways, this edition is like a first edition. Coverage has expanded from being primarily a book with a medical microbiology emphasis to one with a more balanced emphasis of microbiology in general. Following is a summary of the major changes in this edition. Many of the older photos have been replaced with newer ones, and many new photos have been added. Chapter 1 provides an introduction to microbiology and presents a perspective on the places Bacteria and Archaea occupy in the biological world. Chapter 11 covers some of the most important groups within the Domain Bacteria. Chapters 13 and 14 do the same for the Domains Archaea and Eukarya, respectively. The chapters that follow continue the process: The next chapters cover the microbes themselves, beginning with viruses Chapter 10 , and followed by chapters on Domain Bacteria Chapters 11 and 12 , Domain Archaea Chapter 13 , and Domain Eukarya Chapters 14 through The book finishes with chapters on quantitative techniques Chapter 18 , medical, environmental, and food microbiology Chapter 19 , and host defenses Chapter An appendix illustrating major metabolic pathways combined with tables to show reactants and products of each completes this edition. Other topics have been expanded. Cooked Meat Broth was added to Chapter 3 and the anaerobic jar has been updated. Chapter 4 has expanded coverage of electron microscopy. Paraspore crystal stain and cell wall stain have been added to Chapter 6. DNase Test Agar has been expanded in Chapter 7. The Winogradsky column and sulfur cycle have been added to Chapter 19, and the nitrogen cycle has been expanded. Speaking for both Burt and me, we hope you find this edition of the Photographic Atlas for the Microbiology Laboratory more useful than ever.

Chapter 5 : [Microbiology] Atlas of Growth Patterns on Agar | Free Medical Atlas

A Photographic Atlas for the Microbiology Laboratory, Fourth Edition by Michael J. Leboffe and Burton E. Pierce is intended to act as a supplement to introductory microbiology laboratory manuals. This full-color atlas can also be used in conjunction with your own custom laboratory manual.

Chapter 6 : [Microbiology] Types of Microscopy | Free Medical Atlas

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Chapter 7 : A Photographic Atlas For The Microbiology Laboratory by Michael J. Leboffe

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Chapter 8 : Microbiology: A Photographic Atlas for the Laboratory by Steven K. Alexander

The atlas can be used alone but also has been designed to be used in conjunction with Exercises for the Microbiology Laboratory, 4e, by Leboffe & Pierce, with images keyed to specific exercises. A Photographic Atlas for the Microbiology Laboratory, 4e is part of our CustomLab program.

Chapter 9 : Alexander & Strete, Microbiology: A Photographic Atlas for the Laboratory | Pearson

The many reviews about A Photographic Atlas for the MicrobiologyLaboratory before purchasing it in order to gage whether or notit would be worth my time, and all praised APhotographic Atlas for the Microbiology Laboratory, declaringit one of the best, something that all readers will enjoy.